5.30μ (bridge) as well as the bands commonly associated with the triphenylmethyl phosphonium ion. Acidification of III with dilute hydrochloric acid produced immediate decolorization and decaborane was recovered in 50% yield. Air oxidation and hydrolysis of III was not apparent after three months at ambient temperature.

It appears that this material is the triphenylmethylphosphonium salt of decaborane and is the first such material to be isolated as a pure stable

 $(C_6H_5)_3P^+CH_2^- + B_{10}H_{14} \longrightarrow (C_6H_5)_3P^+CH_3B_{10}H_{13}^$ substance.

The reactions of other phosphine methylenes with decaborane are under investigation.

ROHM & HAAS COMPANY

REDSTONE ARSENAL RESEARCH DIVISION HUNTSVILLE, ALABAMA M. FREDERICK HAWTHORNE RECEIVED MAY 23, 1958

ISOLATION OF GUANOSINE DIPHOSPHATE FUCOSE FROM AEROBACTER AEROGENES

Sir:

We wish to report the isolation of a new sugar nucleotide, guanosine diphosphate fucose, from a strain of Aerobacter aerogenes1 that produces a polysaccharide containing L-fucose.²

The nucleotides from 150 g. wet weight of bacteria were extracted with boiling 70% ethanol, precipitated with mercuric ion, and chromatographed on a Dowex 1-Cl⁻ column.³ Elution with 0.01 N HCl and increasing concentrations of NaCl yielded an ultraviolet absorbing peak which was determined to be 75% uridine diphosphate glucose and uridine diphosphate galactose by enzymatic analysis.⁴ This peak contained a total of $10 \ \mu M$. of nucleotides calculated as uridine from spectral data. A minor component of this fraction amounting to 10% of its optical density at 260 m μ could be isolated by paper electrophoresis or paper chromatography. The ultraviolet absorption spectrum of this component was typical of a guanosine derivative. Its mobility during electrophoresis in sodium formate buffer, pH 3.5, was less than guanosine diphosphate, but greater than guanosine monophosphate while it had a higher $R_{\rm f}$ than guanosine monophosphate when chromatographed with ethanol-neutral ammonium acetate solution.⁵

The isolated component was analyzed colorimetrically⁶ and found to contain approximately $0.8 \ \mu M.$ of 6-deoxyhexose per $\mu M.$ guanosine. The absorption peak at 400 m μ given by the guanosine derivative in this test was identical in shape with that given by authentic fucose and also disappeared at the same rate upon dilution with water.⁷

Hydrolysis of the guanosine derivative in 0.01N HCl at 100° for 10 minutes liberated a compound having an $R_{\rm f}$ identical with that of fucose

(1) Strain A₃S₁ (ATCC 12657).

(2) J. F. Wilkinson, W. F. Dudman and G. O. Aspinall, Biochem. J., 59, 446 (1955).

(3) E. Cabib, L. F. Leloir and C. E. Cardini, J. Biol. Chem., 203, 1055 (1953).

(4) H. M. Kalckar, E. P. Anderson, and K. J. Isselbacher, Biochim. Biophys. Acta, 20, 262 (1956).

(5) A. C. Paladini and L. F. Leloir, Biochem. J., 51, 426 (1952).

(6) Z. Dische and L. B. Shettles, J. Biol. Chem., 175, 595 (1948).

(7) Z. Dische and L. B. Shettles, ibid., 192, 579 (1951).

when chromatographed with butanol-acetic acidwater,⁸ phenol-water,⁸ or pyridine-ethyl acetate-water.⁹ In addition, an ultraviolet absorbing compound was formed which exhibited the same chromatographic and spectral properties as guanosine diphosphate. Longer hydrolysis led to the formation of a second ultraviolet absorbing compound which exhibited the same properties as guanosine monophosphate.

In view of the well-established role of the uridine sugar nucleotides as glycosyl donors in the biosynthesis of many complex saccharides, it is an interesting variation to find fucose occurring in a guanosine nucleotide. The only other guanosine sugar nucleotide known at present is guanosine diphosphate mannose.^{10,11}

(8) S. M. Partridge, Biochem. J., 42, 238 (1948).

(9) M. A. Jermyn and F. A. Isherwood, ibid., 44, 402 (1949).

(10) E. Cabib and L. F. Leloir, J. Biol. Chem., 206, 779 (1954).

(11) J. L. Strominger, Biochim. et Biophys. Acta, 17, 283 (1955).

(12) U. S. Public Health Service Postdoctoral Fellow.

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Received April 5, 1958

ORGANOBORON COMPOUNDS. X. MIXED TRIALKYLBORANES DISTILLABLE WITHOUT DISPROPORTIONATION^{1,2}

Sir:

It was suggested recently¹ that mixed trialkylboranes characterized by the presence of a *t*-butyl group may manifest unusual stability to disproportionation. One such substance, diisobutyl-t-butylborane, was described previously and its stability to disproportionation was attributed to steric interference with the disproportionation mechanism.¹

We wish to describe now the first distillable trialkylborane containing three dissimilar alkyl groups, namely, t-butyl-isobutyl-n-amylborane. This substance, b.p. $43.5-44.0^{\circ}$ at 0.5 mm., n^{25} D 1.4296, d^{25} 0.7506, was fractionally distilled twice in vacuo without decomposition, rearrangement or disproportionation. Anal. Calcd. for $C_{13}H_{29}B$: B, 5.52. Found: B, 5.56. MRD: calcd.,³ 67.30; obsd., 67.48. Oxidation with alkaline hydrogen peroxide¹ produced equimolar quantities of *t*-butyl, isobutyl and *n*-amyl alcohols in high yield.

t-Butyl-isobutyl-*n*-amylborane was prepared in two ways: (a) in 50% yield by the alkylation of n-amyldifluoroborane with t-butylmagnesium chloride in anhydrous ether; (b) in 30% yield by the reaction of t-butyl-di-n-amylborane with isobutyl-magnesium bromide.⁴ The physical constants and the infrared spectra of the two samples were practically identical. In connection with method (a), it is noteworthy that one t-butyl group derived from the Grignard reagent rearranges to isobutyl during the alkylation reaction. Concerning

(1) Previous paper, G. F. Hennion, P. A. McCusker and A. J. Rutkowski, THIS JOURNAL, 80, 617 (1958).

(2) Contribution from the Radiation Project operated by the University of Notre Dame and supported in part under Atomic Energy Commission Contract AT-(11)-38.

(3) The B-C bond refraction was taken as 1.93.

(4) S. L. Clark and J. R. Jones, Abstracts, 133rd Meeting, American Chemical Society, San Francisco, April, 1958, p. 34-L.